

Available online at www.sciencedirect.com

SciVerse ScienceDirect

journal homepage: <http://www.elsevier.com/locate/repbio>

Short communication

Ovarian fluid influences sperm performance in lake trout, *Salvelinus namaycush*

Patrick M. Galvano^a, Katelynn Johnson^a, Chris C. Wilson^b, Trevor E. Pitcher^a, Ian A.E. Butts^{a,*}

^aDepartment of Biological Sciences, University of Windsor, 401 Sunset Ave., Windsor, Ontario N9B 3P4, Canada

^bAquatic Research and Development Section, Ontario Ministry of Natural Resources, Trent University, 2140 East Bank Drive, Peterborough, Ontario K9J 7B8, Canada

ARTICLE INFO

Article history:

Received 3 August 2012

Accepted 8 February 2013

Keywords:

Motility

Velocity

Longevity

Reproduction

Aquaculture

CASA

ABSTRACT

The objectives of this study were to determine whether (i) the presence and concentration of ovarian fluid (OF) affects sperm performance traits, and (ii) variation in sperm performance traits is due to male identity, female identity, and/or male × female interactions in lake trout, *Salvelinus namaycush*. Spermatozoa from four males were activated in river water and OF from four females at two concentrations (10 and 15%). Presence of ovarian fluid influenced sperm traits; no differences were detected between 10 and 15% OF. Sperm traits varied depending on parental identity, such that sperm of some males perform better in the OF of all females and that in OF of some females sperm traits are higher for all males.

© 2013 Society for Biology of Reproduction & the Institute of Animal Reproduction and Food Research of Polish Academy of Sciences in Olsztyn. Published by Elsevier Urban & Partner Sp. z o.o. All rights reserved.

1. Introduction

In externally fertilizing teleosts, each female creates a unique spawning environment by simultaneously expelling her ovarian fluid (OF), along with an egg batch [1–3]. Different concentrations of OF have been shown to influence sperm performance traits [4–6] and when activated in OF, sperm performance can depend upon individual male by female interactions, suggesting a possible role in cryptic female choice [3,7,8]. Within this context, ionic [8], biochemical [9], and genetic components (i.e. genes of the major histocompatibility

complex) [10] of the OF have played a key role in stabilizing the micro-environment around the micropyle [1], which in turn has increased the fertilizing ability of sperm [11,12].

Thus, the objectives of this study were to determine whether (i) the presence and concentration of OF affects sperm performance traits, and (ii) variation in sperm performance traits is due to male identity, female identity, and/or male × female interactions in lake trout, *Salvelinus namaycush*. Lake trout spawn in nocturnal aggregations, where several males spawn simultaneously with a female, thus intense sperm competition and gamete selection are probable in the wild [13].

* Corresponding author. Present address: DTU AQUA, National Institute for Aquatic Resources, Section for Marine Ecology, Technical University of Denmark, Charlottenlund Slot, Jægersborg Allé 1, 2920 Charlottenlund, Denmark. Tel.: +45 35 88 33 00; fax: +45 35 88 33 33.

E-mail addresses: ianb@aqu.dtu.dk, iana.e.butts@gmail.com (Ian A.E. Butts).

1642-431X/\$ – see front matter © 2013 Society for Biology of Reproduction & the Institute of Animal Reproduction and Food Research of Polish Academy of Sciences in Olsztyn. Published by Elsevier Urban & Partner Sp. z o.o. All rights reserved.

<http://dx.doi.org/10.1016/j.repbio.2013.02.001>

2. Materials and methods

Sperm traits were measured in lake trout from the Ontario Ministry of Natural Resources Codrington Fisheries Research Facility. The origin of broodstock and rearing conditions prior and during this study are reported in Butts et al. [7]. Gametes and their associated fluids were collected from four males and four females in November 2011, which represents peak spawning [14]. Mean (\pm SEM) fork length and body weight were 684.5 ± 12.3 mm and 4231.3 ± 423.9 g for the males and 606.3 ± 17.9 mm and 2953.3 ± 271.4 g for the females, respectively. Age of these fish ranged from 9 to 11 years. The fish were anaesthetized with MS-222 prior to stripping of gametes. The urogenital pore was wiped dry to avoid contamination. Pressure was then applied to the abdomen and milt was collected using sterilized pipettes. Egg batches, along with the OF, were collected and OF was then separated from the eggs with 1 mm mesh. Gametes were stored in a cooler filled with ice packs (4–6 °C) until further analyses (sperm were activated within 5 h). Sperm performance traits were analyzed at 5, 10, and 15 s post-activation (PA) using methods detailed in Butts et al. [7]. In brief, sperm was activated with inflowing river water at ~ 7.0 °C and pH 7.0. The following sperm traits were evaluated: average path velocity (VAP, hereafter referred to as sperm velocity), linearity, percent motility, and longevity [15].

For each male ($n = 4$), sperm was activated in water (0% OF) and OF from each female ($n = 4$) at 10 and 15% OF concentrations. Two replicate sperm-activations were conducted for each treatment combination; the mean of these two independent activations was used for statistical analyses. We used 10 and 15% OF concentrations in the activation media as it seems likely that sperm would encounter relatively low concentrations of OF during a spawning event, as OF comprises only 5–20% of the total egg volume in lake trout [7]. Data were analyzed using one-way mixed-model ANOVAs. The independent variable was OF concentration (fixed effect), and male was considered a random factor. Models were run separately at each PA time using SAS statistical analysis software (v.9.1; SAS Institute Inc., Cary, NC, USA).

Furthermore, sperm from each male was activated in each female's OF (four males \times four females \times two replicate sperm-activations; the mean of the two independent activations was used for statistical analyses). No significant differences in sperm performance traits were detected between 10 and 15% OF concentrations (see below); thus sperm were therefore activated in 10% OF. Sperm performance traits were analyzed using a factorial ANOVA containing the male (random effect) and female (random effect) main effects as well as the male \times female interaction. Models were run separately at each PA time.

3. Results and discussion

The presence of OF had a significant effect on sperm velocity at 5 ($p < 0.01$), 10 ($p < 0.05$), and 15 s ($p < 0.05$) PA; sperm motility at 5 ($p < 0.001$), 10 ($p < 0.01$), and 15 s ($p < 0.01$) PA; and linearity at 10 ($p < 0.05$), and 15 s PA ($p < 0.05$) (Fig. 1). Moreover, the presence of OF had an effect on longevity

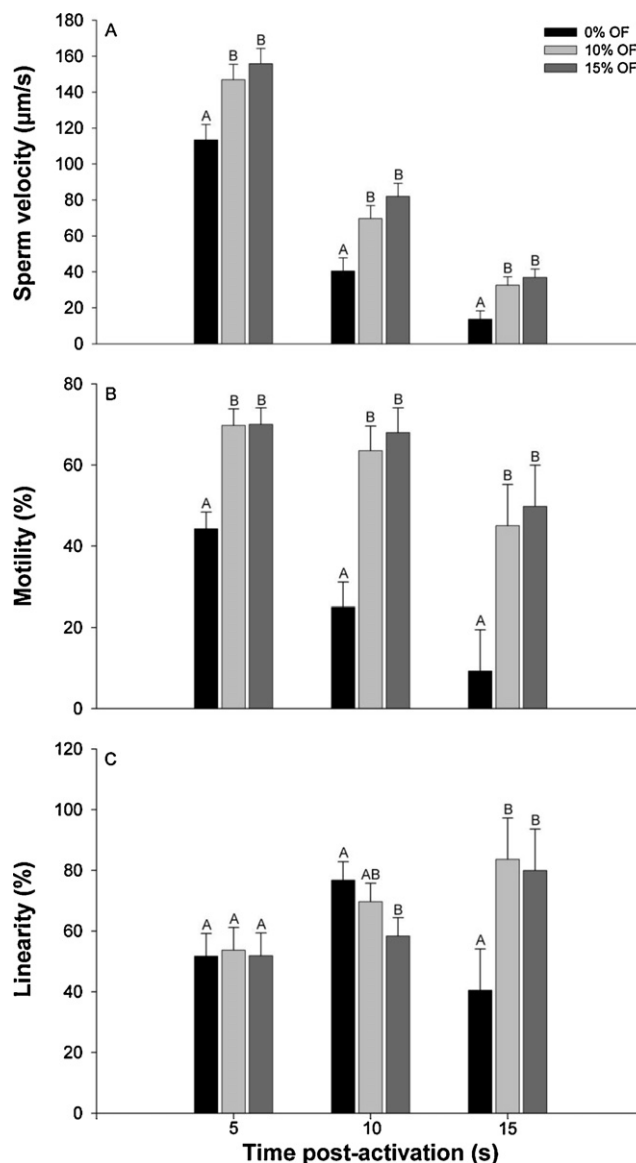


Fig. 1 – The effect of ovarian fluid concentrations (0, 10 and 15%) on average path velocity (A), percent motility (B), and linearity (C) at 5, 10, and 15 s post-activation of sperm in lake trout, *Salvelinus namaycush*. Error bars represent standard errors. Bars without a common superscript differed significantly.

($p < 0.001$), such that sperm swam longer at the 10% (17.78 ± 0.55 s), and 15% OF concentrations (18.84 ± 0.55 s), when compared to 0% OF (11.00 ± 0.79 s). For all of the sperm performance traits, no significant difference was found between the 10% and 15% OF concentrations. Overall, these results suggest that sperm performance traits are enhanced in the presence of OF and that these effects represent OF concentrations at realistic spawning levels. A similar effect has been previously demonstrated in fishes, i.e. sperm swimming speed increased in OF when compared to sperm activated in water alone [4,5]. This has implications for relative reproductive success among males, as sperm velocity and

motility (among other performance traits) has been shown to correlate with competitive fertilization success in fishes [15,16]. Thus, our results further strengthen the importance of incorporating OF into sperm-activation and fertilization media to represent more natural spawning environments, particularly if cryptic female choice is a possibility.

For all sperm performance traits there were non-significant male × female interactions ($p > 0.05$), therefore male and female main effects were interpreted. For sperm velocity there was a significant male effect at 5 ($p < 0.05$), and 10 s PA ($p < 0.05$; Fig. 2A), and a significant female effect at 15 s PA ($p < 0.01$; Fig. 2B), while for sperm motility a significant male effect was detected at 15 s PA ($p < 0.01$; Fig. 2C). A

significant male effect was detected for linearity at 5 ($p < 0.01$), and 10 s PA ($p < 0.001$; Fig. 2E), and a significant female effect was detected at 15 s PA ($p < 0.05$; Fig. 2F). Ovarian fluid from the different females affected longevity ($p < 0.01$), while the male effect was non-significant. Together, these findings suggest that some males perform better in the OF of all females and that some females have OF in which sperm traits are higher for all males. On the contrary, we found no interaction effect and therefore no evidence of a potential mechanism of cryptic female choice (male × female interaction) as detected in other fish species [3,8]. Thus, further investigation is required to understand the mechanisms by which OF influences sperm performance in lake

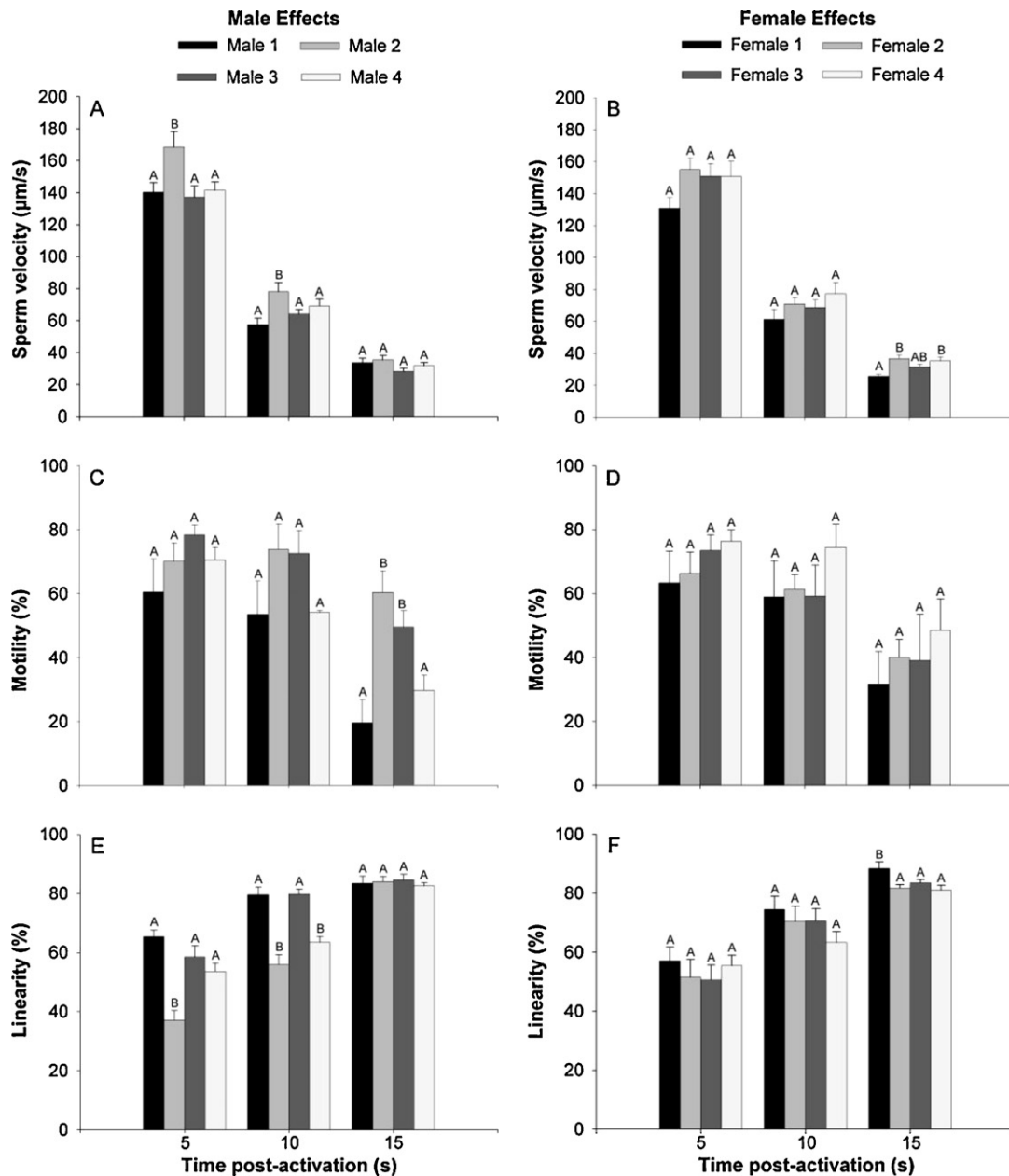


Fig. 2 – The effects of male identity, female identity, and the male × female interaction on average path velocity, percent motility, and linearity in lake trout, *Salvelinus namaycush*. Sperm traits were activated in 10% ovarian fluid. For all sperm traits there were non-significant male × female interactions, therefore the male (A, C, and E) and female (B, D, and F) main effects were interpreted. Error bars represent standard errors. Bars without a common superscript differed significantly.

trout. For instance, it has been shown that the ionic composition of OF enhances sperm performance in Chinook salmon, *Oncorhynchus tshawytscha* [8], while sperm-activating proteins enhance sperm performance in Pacific herring, *Clupea pallasii* [9]. Future studies should also investigate how OF impacts fertility as it has been suggested that sperm released closer to an egg will be exposed to higher concentrations of OF and possibly gain a fertilization advantage [5].

Acknowledgements

Funding for this study was provided by the Natural Science and Engineering Research Council of Canada, Ontario Ministry of Innovation and Canada Foundation for Innovation (to TEP) and an Ontario Ministry of Research and Innovation Postdoctoral Fellowship (to IAEB). Logistic support was provided by the Ontario MNR Codrington Fisheries Research Facility. Thanks to Bill Sloan and Scott Ferguson for rearing fish and aiding with gamete collection.

REFERENCES

- [1] Lahnsteiner F. The influence of ovarian fluid on the gamete physiology in the Salmonidae. *Fish Physiology and Biochemistry* 2002;27:49–59.
- [2] Rosengrave P, Gemmell NJ, Metcalf V, McBride K, Montgomerie R. A mechanism for cryptic female choice in chinook salmon. *Behavioural Ecology* 2008;19:1179–85.
- [3] Urbach D, Folstad I, Rudolfson G. Effects of ovarian fluid on sperm velocity in Arctic charr (*Salvelinus alpinus*). *Behavioural Ecology and Sociobiology* 2005;57:438–44.
- [4] Dietrich GJ, Wojtczak M, Slowinska M, Dobosz S, Kuzminski K, Ciereszko A. Effects of ovarian fluid on motility characteristics of rainbow trout (*Oncorhynchus mykiss* Walbaum) spermatozoa. *Journal of Applied Ichthyology* 2008;24:503–7.
- [5] Turner E, Montgomerie R. Ovarian fluid enhances sperm movement in Arctic charr. *Journal of Fish Biology* 2002;60:1570–9.
- [6] Wojtczak M, Dietrich GJ, Slowinska M, Dobosz S, Kuzminski H, Ciereszko A. Ovarian fluid pH enhances motility parameters of rainbow trout (*Oncorhynchus mykiss*) spermatozoa. *Aquaculture* 2007;270:259–64.
- [7] Butts IAE, Johnson K, Wilson C, Pitcher TE. Ovarian fluid enhances sperm velocity based on relatedness in lake trout, *Salvelinus namaycush*. *Theriogenology* 2012;78:2105–9.
- [8] Rosengrave P, Taylor H, Montgomerie R, Metcalf V, McBride K, Gemmell NJ. Chemical composition of seminal and ovarian fluids of chinook salmon (*Oncorhynchus tshawytscha*) and their effects on sperm motility traits. *Comparative Biochemistry and Physiology Part A* 2008;152:123–9.
- [9] Oda S, Igarashi Y, Ohtake H, Sakai K, Shimizu N, Morisawa M. Sperm-activating proteins from unfertilized eggs of the pacific herring *Clupea pallasii*. *Development Growth and Differentiation* 1995;37:257–61.
- [10] Gasparini C, Pilaastro A. Cryptic female preference for genetically unrelated males is mediated by ovarian fluid in the guppy. *Proceedings of the Royal Society of London B Biological Sciences* 2011;278:2495–501.
- [11] Billard R. Effects of coelomic and seminal fluids and various saline diluents on the fertilizing ability of spermatozoa in the rainbow trout, *Salmo gairdneri*. *Journal of Reproduction Fertility* 1983;68:77–84.
- [12] Hatef A, Niksirat H, Alavi SMH. Composition of ovarian fluid in endangered Caspian brown trout, *Salmo trutta caspius*, and its effects on spermatozoa motility and fertilizing ability compared to freshwater and a saline medium. *Fish Physiology and Biochemistry* 2009;35:695–700.
- [13] Esteve M, McLennan DA, Gunn JM. Lake trout (*Salvelinus namaycush*) spawning behaviour: the evolution of a new female strategy. *Environmental Biology of Fishes* 2008;83:69–76.
- [14] Johnson K, Butts IAE, Wilson C, Pitcher TE. Sperm quality of hatchery-reared lake trout *Salvelinus namaycush* throughout the spawning season. *North American Journal of Aquaculture* 2013;75:102–8.
- [15] Gage MJG, Macfarlane CP, Yeates S, Ward RG, Searle JB, Parker GA. Spermatozoal traits and sperm competition in Atlantic salmon: relative sperm velocity is the primary determinant of fertilization success. *Current Biology* 2004;14:44–7.
- [16] Linhart O, Rodina M, Gela D, Kocour M, Vandeputte M. Spermatozoal competition in common carp (*Cyprinus carpio*): what is the primary determinant of competition success? *Reproduction* 2005;130:705–11.